IMMUNOLOGY IN THE CLINIC: DIABETES UPDATE

Series Editor: Mark Peakman

# Clinical and Experimental Immunology

# Progress in immune-based therapies for type 1 diabetes

OTHER ARTICLES PUBLISHED IN THIS SERIES

Immunological biomarkers: Catalysts for translational advances in autoimmune diabetes. Clinical and Experimental Immunology 2013, 172: 178-85.

# M. von Herrath,\* M. Peakman<sup>†</sup> and B. Roep<sup>‡</sup>

\*Center for Type 1 Diabetes Research, La Jolla Institute for Allergy and Immunology, La Jolla, CA, USA, †Department of Immunobiology, School of Medicine, King's College London, London, UK, and †Department of Immunohematology and Blood Transfusion, Leiden University Medical Center, Leiden, the Netherlands

Accepted for publication 30 January 2013 Correspondence: M. Peakman, Department of Immunobiology, King's College London School of Medicine, 2nd Floor, Borough Wing, Guy's Hospital, London SE1 9RT, UK. E-mail: mark.peakman@kcl.ac.uk

All authors contributed equally to this article.

# **Summary**

Immune-based therapies that prevent type 1 diabetes or preserve metabolic function remaining at diagnosis have become a major objective for funding agencies and international trial consortia, and receive backing from notable patient advocate groups. The development of immune-based therapeutic strategies in this arena requires a careful balancing of the risks of the therapy against the potential benefits, because many individuals are diagnosed or identified as being at increased risk of disease in early childhood, a period when manipulation of the developing immune system should be undertaken with caution. In addition, a therapy exists (daily insulin injection) that is lifesaving in the acute stages of disease and can be used effectively over a lifetime as maintenance. Conversely, the disease is increasing in incidence; is peaking in ever-younger age groups; carries significant risk of increased morbidity and early mortality; and remains difficult to manage effectively in many settings. With these issues in mind, in this article we review progress towards immune-based strategies for this chronic autoimmune disease.

**Keywords:** autoimmunity, diabetes, immunotherapy

# The perspective

With the exception of one or two early attempts at disease modulation, the field of immunotherapy for type 1 diabetes did not develop significant momentum until the 1980s, during which a series of studies were initiated that made use of a drug (cyclosporin) which had, by then, revolutionized immune suppression in the setting of organ transplantation. Some 20 years on from those early successes, in 2007 we reviewed the status of intervention and prevention trials for type 1 diabetes [1]. The timing of our commentary was significant; the first major advance since cyclosporin had recently emerged, notably with the publication of two studies using monoclonal antibodies (mAbs) targeting CD3 and engineered to have limited Fc binding, both of which demonstrated clinically relevant efficacy with manageable toxicity [2,3]. At that stage we discussed the fact that these drugs (subsequently emerging as teplizumab and otelixizumab) were lead agents at the head of a therapeutic pipeline of immunomodulators. These included several drugs that were emerging from the fields of transplantation immunology and as treatments for other autoimmune and inflammatory diseases, as well as disease-specific, antigen-based therapeutics. In a subsequent, related review paper we highlighted the potential and pitfalls of harnessing these agents into combinations [4], including a proposed 'designer combo' of anti-inflammatory + immune modulator + antigen. Moreover, to facilitate the pipeline, during the same period significant infrastructure was emerging in the form of clinical trial networks, within which clinical studies could be conducted to agreed and standardized designs and protocols. The exemplar of this approach is Type 1 Diabetes TrialNet (http://www.diabetestrialnet.org). There was even significant and demonstrable interest in this disease space being displayed by large pharmaceutical concerns. Consequently, as a result of this constellation of events, in 2007 the clinical trial horizon for type 1 diabetes was viewed with the expectation of success and progress. Some 6 years on, several key questions emerge. What has become of the pipeline and the combination approaches? Using the same format as the 2007 paper, we have updated the data tables with new or contemporary information on trials conducted or in progress at that time, and added information on new and ongoing studies. Information-gathering is based largely

on the US National Institutes of Health-sponsored website ClinicalTrials.gov (http://www.clinicaltrials.gov) and the European equivalent (EU Clinical Trials Register; https:// www.clinicaltrialsregister.eu/index.html), as well as our knowledge of the sector. Our analyses include studies conducted in the predisease setting, before diabetes onset, for both antigen-specific and non-antigen-specific approaches [primary (high genetic risk) and secondary (high risk identified by islet cell autoantibody positivity) prevention studies, Tables 1 and 2, respectively] and trials in which recruitment centres on subjects who have already developed disease (intervention studies; Tables 3 and 4, respectively). There is a further update on trials using combination approaches (Table 5). What have we learned from the clinical trials that have been conducted? Has our general understanding of the disease altered in any respect in the intervening period, such that we might review our therapeutic options?

### Trial design for intervention studies

With the premise that type 1 diabetes is an immunemediated disorder, most efforts to intervene in disease pathogenesis involve immune-based therapy. Without exception, primary study end-points tend to focus on preservation of  $\beta$  cell function, as measured by stimulated C-peptide production after a standardized food challenge (oral glucose tolerance test, OGTT) or glucagon injection. This is a justifiable criterion that is accepted by regulatory agencies such as the US Food and Drug Administration and European Medicines Agency. Several clinical trials assessing immune interventions (teplizumab, otelixizumab, rituximab, abatacept; see Table 4) show a temporary delay in the loss of  $\beta$  cell function as defined by OGTT, while injection of a heat shock protein-derived peptide (DiaPep277; Table 2) only showed a beneficial effect based on glucagonstimulated measurement of  $\beta$  cell function, but not on OGTT; the reasons for this intriguing finding are not yet known. Improved glycaemic control, as measured by reduction in glycated haemoglobin levels (HbA1c), should not be considered a useful end-point going forward, even though it was used (albeit unsuccessfully) in the Phase III teplizumab (anti-CD3) trial. Patients enrolled into intervention trials should be treated to prespecified HbA1c target levels using standard clinical care, and thus any differences between treatment and placebo groups raise concerns about study design and conduct. In general, therefore, changes in immune correlates of the autoimmune process [5] have not been selected as study end-points, even though the disease process is immune-mediated. Given that defining changes in disease progression by C-peptide measurement imposes long-term study follow-up, and new insights which suggest that  $\beta$  cell function does not necessarily equate with  $\beta$  cell mass [6], there is a strong argument to be made that the field should shift towards alternative, immune-based endpoints that can deliver more rapidly and potentially in smaller-sized treatment groups, at least at a 'proof-of-concept' stage [5,7].

As the unmet medical needs and potential benefits of successful immunotherapy are greatest in children, it is evident that the inclusion of children in clinical trials is highly desirable, provided that there is adequate risk assessment. Indeed, the inclusion of younger patients in the rituximab trial secured short-term efficacy that would have remained unnoticed if subjects only beyond 18 years of age had been recruited [8]. Effects of otelixizumab in older patients became apparent only upon extended follow-up [9]. In addition to age, the timing of inclusion and window of opportunity for success in relation to disease progression remain poorly defined. Depending on the type of intervention, it may prove difficult to treat during the medical emergency of newly manifested disease, although early enrolment (typically 3 months after diagnosis) has become the common inclusion criterion for intervention trials. As  $\beta$ cells survive up to decades after diagnosis, together with insulitic lesions [10,11], there is in reality no reason to exclude patients beyond 3-6 months after diagnosis who have measurable C-peptide, other than the slower slope in decline of stimulated  $\beta$  cell function and associated reduced statistical power to define treatment-induced changes. This, again, argues for alternative (surrogate) end-points of therapeutic efficacy [5]. Intervention studies beyond the first year after onset would also avoid the confounding effect of the natural remission and temporarily reduced insulin needs (known as the 'honeymoon') that often occurs shortly after initiation of insulin replacement therapy. In terms of staging of patients during stratification in trial enrolment, we may need to take lessons from new insights emerging from studies on disease tissue (via the Network for Pancreatic Organ Donors with Diabetes; nPOD [10]) and Phase III clinical trials failing to reach end-points [12,13]. Both of these imply that type 1 diabetes may be a very heterogeneous disease, manifesting differently in different patient groups and geographical locations. An intriguing example is that of abatacept, which appeared to worsen clinical outcome in African American subjects [14]. In addition, the average age at disease onset of patients enrolled on the Indian subcontinent into the teplizumab Phase III study was 44 years [13], an age of disease onset that would usually be considered at the very upper limit. With the exception of oral insulin [15] and proinsulin peptide immunotherapy [16], immunological parameters have not generally been used in selection or randomization of patients in clinical trials. Lessons from the islet transplantation setting, in which baseline immune correlates determine clinical outcome [17–19], may be of use here and it is conceivable that incorporating immune correlates into trial design may improve the chance of detecting therapeutic efficacy and indicate subpopulations of patients with particular benefit, lack of efficacy or even adverse responses

 Table 1.
 Completed, ongoing and planned prevention trials in type 1 diabetes (T1D) using antigen-specific approaches.

Agent	Stage of development in 2007	Details (including ClinicalTrials.gov Identifier)	References and links	
Parenteral insulin Parenteral insulin	Pilot, completed 1993 Pilot, completed 1998	Small, pilot study, suggestive of efficacy Small, pilot study, suggestive of efficacy	[61] [62]	
Parenteral insulin (DPT-1)	Large efficacy study, completed 2002	No effect seen on disease progression	[63] and http://www. diabetestrialnet.org	
Oral insulin (DPT-1)	Large efficacy study, completed 2005	No effect seen on disease progression; however, strong evidence from subanalysis of significant treatment effect on subjects with strong evidence of insulin autoimmunity. Repeat study planned (NCT00004984)	[15] and http://www. diabetestrialnet.org	
Intranasal insulin (INIT I)	Phase I, completed 2004	No acceleration of loss of beta cell function in individuals at risk for T1D. Immune changes consistent with mucosal tolerance to insulin detected	[64]	
				Progress to date
Intranasal insulin (DIPP)	Ongoing	RPCT trial of daily intranasal short-acting human insulin (1 unit/kg) in at-risk (high-risk HLA, dual autoantibody-positive) (NCT00223613)	[65] and http://research. utu.fi/dipp	Study published in 2008 [28]; no effect on primary end-point of progression to Type 1 diabetes
Intranasal insulin (INIT II)	Phase II, to start 2006/07	RPCT of intranasal insulin in at-risk (autoantibody positive) relatives (NCT00336674)	https://studies. thegeorgeinstitute. org/init/PMID	Multi-centre Phase II started 2008 in Australia and extended to Munich in 2011. Currently ongoing. Recent data show significant treatment-associated blunting of insulin antibody resonae [66]
Oral insulin	Efficacy study, starts	Repeat of oral arm of DPT-1. Randomized, double blind, nacelyo controlled trial of oral insulin in	http://www. diahetestrialnet org	Started recruitment in 2007 with > 250 enrolled by end 2012. Reporting possible in 2014/2015
	70,700.7	at-risk (autoantibody positive) relatives with insulin autoantibodies as inclusion criterion (NCT00419562)	ulabetestijaniet.org	oy cita 2012. Nepotting possible in 2014/2015
Oral insulin	Pilot, starts 2006/07	Pre-POINT study: dose finding in children with high genetic risk for type 1 diabetes  EudraCT number: 2005-001621-29	http://www.diabetes- point.org/	Ongoing [67]
				New studies: progress to date
Injection (s.c.) GAD-Alum (DiAPREV-IT)	n.a.	Study designated as ongoing but not recruiting on ClinicalTrials.gov. (NCT01122446). Last subject enrolled January 2012. Planned as double-blind, randomized study to determine the safety and the effect of Diamyd* on the progression to type I diabetes in children with multiple islet cell	http://www.diamyd.com/ docs/trialsDiabetes. aspx?section=trials	Ongoing
		autoantiboules		

RPCT: randomized placebo-controlled trial; DPT-1: diabetes prevention trial-1; POINT: Primary Oral Insulin Trial; DiAPREV-IT: Diabetes Prevention Immune-Tolerance. HLA: human leucocyte antigen; n.a.: not available; s.c.: subcutaneous;

Table 2. Completed, ongoing and planned prevention trials in type 1 diabetes (T1D) using non-antigen-specific approaches.

Agent	Stage of development in 2007	Details (including ClinicalTrials.gov Identifier)	References and links	
Ketotifen (histamine antagonist)	Pilot. completed 1994	No effect	[89]	
CvA	Pilot, completed 1996	Delay but not prevention in high-risk group	[69]	
Nicotinamide (Deutsche Nicotinamide	Efficacy study, completed	No effect	[20]	
Intervention Study; DENIS)	1998			
Nicotinamide European Nicotinamide	Efficacy study, completed	No effect	[71]	
Diabetes Intervention Trial (ENDIT)	2004			
Various combinations nicotinamide, CyA, insulin, Vit E	Pilots 1 <i>9</i> 94–2005	No additive effects	[72–74]	
BCG	Various pilots	No effect	[75–77]	
Gluten-free diet	Pilot, completed 2002	No effect on autoantibodies or disease	[78]	
				Progress to date
Vitamin D3	Phase I, ongoing	Pilot two-arm RCT to study feasibility of 2000 IU per day of vitamin D for the primary prevention of Type		Completed [79]; demonstrated feasibility and safety
		1 Diabetes (main objective to compare 2000 IU with 400 IU (standard of care) in terms of safety and		
		vitamin D-related measurements and feasibility)		
		(NCT00141986)		
Hydrolyzed cow's milk (TRIGR)	Phase I, ongoing	Primary Prevention Study for Type 1 Diabetes in	http://trigr.epi.	Recruitment completed. Finnish
		Children at Risk. RCT with assignment to hydrolysed	usf.edu/.[80].	pilot component (NCT00570102)
		(NCT00179777, NCT00570102)		cell autoantibodies [81]
Docosahexaenoic acid (DHA); omega-3	Pilot, ongoing	NIP study - Nutritional Intervention to Prevent	http://www.	In follow-up [82,83]
fatty acids		Diabetes. Pilot and feasibility study of DHA	diabetestrialnet.org	
		supplementation anti-inflammatory effects during		
		late pregnancy or after birth in high-risk infants (NCT00333554)		
				New studies: progress to date
Removal of Bovine Insulin From Cow's	Pilot	Finnish Dietary Intervention Trial for the Prevention of		Completed. Bovine insulin-free CMF
Milk (FINDIA) Pilot Study		Type 1 Diabetes. Primary prevention pilot study of		reduced the cumulative incidence
		weaning high-risk genotype infants to a bovine		of autoantibodies by age 3 years in
		insulin-free cow's milk formula (CMF) with islet cell		children at genetic risk of type 1
		autoantibodies as outcome (NCT01055080)		diabetes mellitus [84]
BABYDIET, gluten-free diet in infancy		Randomized open-label primary prevention study of		No significant differences in
		effect of early or late first gluten exposure on islet		autoantibody or diabetes
		cell autoantibody development at 3 years of age in FDRs at high genetic risk (NCT01115621)		development observed [85]
Anti-CD3 mAb (Teplizumab)	Phase II	RPCT to prevent or delay the onset of type 1 diabetes	http://www.	In recruitment
		in FDRs with multiple islet cell autoantibodies and immaired olucose tolerance (NCT01030861)	diabetestrialnet.org	
Abatacept (CTLA-4-Ig)	Phase II	RPCT in autoantibody-positive FDRs to prevent	http://www.	Opening 2013
(8)	11100 11	development of abnormal glucose tolerance	diabetestrialnet.org	
		,		

CyA: cyclosporin A; VitE: vitamin E; BCG: bacille Calmette-Guérin; TRIGR: Trial to Reduce Insulin Dependent Diabetes in the Genetically at Risk; FDR: first-degree relative; RPCT: randomized placebo-controlled trial; mAb: monoclonal antibody; CTLA-4-1g. cytotoxic T lymphocyte antigen 4-immunoglobulin.

Table 3. Completed, ongoing and planned intervention trials in type 1 diabetes (T1D) using antigen-specific approaches.

Injection (s.c.) APL of insulin B chain peptide Injection (s.c.) DiaPep277 Ph (hsp60 peptide)				
	Phase I, completed	NBI-6024 (NCT00873561)	[98]	
	Phase II. completed	Phase II in adults reports preservation of C-peptide	[87.88]	
		at 12–18 months.		
Injection (e.g.)	Dhasa I commisted in IADA	Phase II in children reports no treatment effect	[80] http://www.diamoid.com	
	iase i, completed in LADA	awaited (see below)	[62] IIItp://www.uiaiiiyu.coiii	
				Progress to date
insulin B	Phase I, completed	(NCT00057499)	http://www.immunetolerance.	Study drug safe; induces humoral and
chain in 1FA			org/researcn/autoimmune/ trials/orban1.html	cellular anti-insulin responses; CD4 1 cell have regulatory phenotype [31]
	Phase Ia (ongoing)	Open-label safety and dosing study in	http://www.dvdc.org	Completed; study drug safe; induces
(C19-A3) peptide		long-standing Type 1 diabetes		anti-peptide IL-10 <sup>+</sup> CD4 T cells [16]
Injection (i.m.) PI- DNA Ph	Phase I planned	BHT-3021 – randomized double-blind	http://www.bayhilltx.com/	Completed; report pending
vaccine		placebo-controlled safety and pharmacodynamic study with open-label cross-over (NCT00453375)		
Injection (s.c.) Ph	Phase II	RPCT of GAD-alum in children and adolescents	http://www.diamyd.com	Completed; fasting C-peptide declined from
GAD-Alum		with recent onset Type 1 diabetes. (NCT00435981)		baseline significantly less in patients treated closest to diagnosis compared with the placebo group [29]
				New studies: progress to date
PI (C19-A3) peptide Ph	Phase Ib	RPCT of safety and dose frequency study in new-onset Type 1 diabetes (NCT01536431)	http://www.dvdc.org	Ongoing
GAD-Alum Ph	Phase II	RPCT of effects of GAD-alum on disease	http://www.diabetestrialnet.org	No effect [30]
3	completed	progression in new onset subjects (NCT00529399)		
GAD-Alum Ph	Phase III	RPCT of effects of GAD-alum on disease	http://www.diamyd.com	No effect at 15 months in EU study; both
ter	terminated	progression in new onset subjects. EU (NCT00723411) and USA DIAPREVENT (NCT00751842)		trials terminated [12]
DiaPep277 (hsp60 Ph peptide)	Phase III	RPCT of effects of DiaPep277 in newly diagnosed type 1 diabetes (DIA-AID) (NCT00615264) and	http://www.andromedabio.com	DIA-AID is completed and has reported preservation of C-peptide in response to
		DIA-AID2 (NC101103284) with open-label extension also recruiting (NCT01460251)		a glucagon-stimulated test [90]; DIA-AID2 (using MMTT) is ongoing

APL: altered peptide ligand; IFA: incomplete Freund's adjuvant; PI: proinsulin; MMTT: mixed meal tolerance test; RPCT: randomized placebo-controlled trial; i.m.: intramuscular; i.d.: intradermal; s.c.: subcutaneous; LADA: latent autoimmune diabetes in adult.

Table 4. Completed, ongoing and planned intervention trials in type 1 diabetes (T1D) using non-antigen-specific approaches.

Agent/study title	Stage of development	Details (including Clinical Trials.gov Identifier)	References and links	
Cyclosporin  Nicotinamide Anti-thymocyte globulin plus prednisolone Bacille Calmette–Guérin (BCG) Dizzoxide IFN-α Anti-CD3 mAb hOKT3g1(Ala-Ala); drug subsequently known as Teplizumabla.	Various trials completed 1984–96 Pilot Pilot Pilot Pilot Phase II Phase II Phase III, completed 2002	Remission induced successfully in recent-onset patients, but therapy typically suspended due to unacceptable side effects. No effect. Reduced insulin requirements more than 100 days after therapy; complicated by severe, transient thrombocytopenia. No effect. No effect. Ingested; small pilot; possible effect (NCT00005665). Disease remission out to 18 months.	[91,92] [93] [94] [95–97] [100] [2]	
Anti-CJ5 mAb ChAglyCJ3(TRX4); drug subsequently known as Otelixizumab	Phase II, completed 2005	Keduced Insulin requirement out to 18 months	[6]	Progress to date
PRODIAB (oral protease) Teplizumab (anti-CD3 mAb) treatment of recent-onset type 1 diabetes	Phase I Phase II	No effect on disease or serum cytokines Participants had type 1 diabetes for 4–12 months before treatment with teplizumab or placebo (NCT00378508)		Data published 2009 [101] Decline in insulin secretion reduced but less of an effect than seen in new-onset period. CD8 biomarker in clinical resonnders [102]
Protégé: Teplizumab in new onset type 1 diabetes	Phase III	RPCT in North America, Europe, Israel and India of standard, Iow and partial dose of anti-CD3 mAb at baseline and 26 weeks (NCT00385697)		Primary composite outcome (% patients on <0.5 U/kg per day insulin HbAIC < 6.5% at 1 year not reached, but 5% of teplizumab groups were not taking insulin at 1 year, compared with no patients in the placebo group (P = 0.03) [13]. The Protee's study is still in follow-up
Teplizumab in recently diagnosed type 1 diabetes (AbATE)	Phase II	RPCT in new-onset type 1 diabetes; patients on active drug receive drug at study entry and at 12 months (NCT00129259)	http://www.abatetrial.org/	Enrolment complete; results awaited
Otelixizumab (anti-CD3 mAb) for adults with newly diagnosed type 1 diabetes (DEFEND-1)	Phase III	Randomized placebo-controlled study in new-onset T1D; single dose; primary outcome C-peptide release after mixed meal (NCT00451321; NCT00678886). NCT01123083 (DEFEND-2, follow-up Phase III) and NCT01222078 (redosing study) terminated	http://us.gsk.com/html/media- news/pressreleases/2011/2011_ pressrelease_10039.htm	Study results not yet posted; press release indicates failure to reach primary end-point
Completed extension of Phase II therapeutic trial [3] of Teplizumab		4-year metabolic outcome study (NCT00627146)		Treatment can suppress the rise in insulin requirements of recent-onset type 1 diabetic patients over 48 months, depending on their age and initial residual beta cell function [9]
Anti-CD20 mAb (Rituximab)	Phase II	RPCT in newly diagnosed type 1 diabetes of rituximab on days 1, 8, 15 and 22 (NCT00279305)	http://www.diabetestrialnet.org	The primary outcome at 1 year (residual C-peptide after mixed- meal) was significantly higher in the rituximab than in the placebo group; the rituximab group also had significantly lower levels of HbAI cand recuired less insulin [8]
Autologous umbilical cord blood cells	Phase I	Patients with type 1 diabetes received a single intravenous infusion of autologous umbilical cord blood cells (NCT00305344)		There were no infusion-related adverse events and no evidence of C-peptide preservation at 2 years. An increase in T <sub>reps</sub> and mave T <sub>reps</sub> were observed but the study lacked a control eroup [103,104]
Polyclonal anti-T-lymphocyte globulin (ATG) in type 1 diabetes		Polyclonal rabbit ATG in patients with type 1 diabetes within 4 weeks of diagnosis as bolus of 9 mg/kg followed by 3 consecutive doses of 3 mg/kg (NCT00190502)		Interim analysis reported on 11 subjects in 2004; at 12 months significant reduction in insulin dose and improved stimulated C-peptide levels in the ATG group [105]
Study of thymoglobulin to arrest newly diagnosed type 1 diabetes (START)	Phase II	Study completed 2012 (NCT00515099)	http://www.immunetolerance.org/ studies/study-thymoglobulin- arrest-type-1-diabetes-start	Full study report awaited; reported abstract suggests no effect on T1D progression [22].
Campath 1H* (anti-CD52 antibody; Alemtuzumab)	Phases I/II	Study withdrawn (NCT00214214)		
Autologous dendritic cell (DC) therapy for type 1 diabetes suppression	Phase I	Autologous DCs manipulated using anti-sense oligonudeotides for CD40, CD80 and CD86 and readministered (NCT00445913)		Well tolerated, significant increase in the frequency of B220 + CD11c-B cells observed. Study reported 2011 [106]
Anakinra (recombinant IL-1 receptor antagonist) in newly diagnosed type 1 diabetes	Phase I/II	Exploratory, open-label study of daily anakinra administered for 28 days to 15 children diagnosed <1 week; no effect on blood gene expression profile in vivo; reduced insulin requirements at 1 and 4 months; no effect on C-peptide (NCT00645840)		[107]

Anti-interieukin-1 in diabetes action (AIDA) Canakinumab (anti IL-1β) in Newly Diagnosed	Phase II Phase II	RPCT of anakinra in new onset-type 1 diabetes (NCT00711503) RPCT in new-onset type 1 diabetes (NCT00947427)		Study report awaited; reported abstracts suggest no effect on disease progression [108] Study report awaited; reported abstracts suggest no effect on
diabetes  Beta cell rescue in new onset type 1 diabetes with  Ffalizumah (anti-CD11 at RRTF)	Phase II	Drug withdrawn due to safety concerns (NCT00737763)		disease progression [109] Terminated
Alphaneo (and Control Alphaneo) in recent-onset	Phase II	Open-label; recent-onset patients (< 5 years with residual		In recruitment
type 1 undeeds Alpha-1 antitrypsin in new onset T1D (Glassia*)	Phases I/II	(NCT01304537)	http://www.kamada.com/	Study report awaited
Intravenous CTLA-4-lg in recent onset type 1 diabetes mellitus	Phase II	RPCT shows beneficial effect on C-peptide preservation in new-onser T1D (NCT0050375)		[14]
Etanercept (anti-TNF- $\alpha$ ) in new-onset type 1 diabetes	Phases I/II	RPCT in 18 new-onset T1D patients age 3–18 years (NCT00730392)		Significant retention of C-peptide and lower HbA1C and insulin dose at week 24 in treated group; drug well tolerated [24]; extension studies suspended after FbA warming on sefery (increased riek of lownhown) in Arithen
Calcitriol in new-onset type 1 diabetes Dose-effect relationship of low-dose IL-2 in type 1 diabetes (DF-IL2)	Phases I/II	RPCT (NCT00960635) RPCT in recent-onset T1D (< 2 years) studying effect of repeated administration of low-dose IL-2 on the kinetics of T - (NCT01353833).		No beneficial effect [110] Study report awaited
Autologous haematopoietic stem cell transplantation (AHSCT) for early-onset type 1	Phases I/II	Jegs (XCIO) 200000 Open-label; patients with T1D received AHSCT after pretreatment with cyclophosphamide and ATG		Complete remission, defined as insulin independence, was observed in 15/28 patients and especially in non-DKA
Diabetes intervention with Atorvastatin	Phase II	Multi-centre RPC in 89 patients with newly diagnosed 71D of		Paucins [111] Fasting and stimulated C-peptide levels were not significantly afficient between the second of the s
Interferon-α for diabetes mellitus type 1	Phase II	No ring day adovasami for 10 months (ACL1097/4749) RPCT in new-onset T1D (< 6 weeks) who received ingested hrIFN-cr at 5000 or 30 000 units or placebo once daily for 1		Treatment was safe; low-dose group maintained more $\beta$ cell function 1 year after study enrolment than placebo group
Prevention of Diabetes progression trial (PDPT)		year (NCL100024918) Open-label, single-dose anti-CD25 mAb in new-onset (< 3 months) T1D (NCT100198146)		[114] Completed; data not reported
BCG administration to alter T-lymphocyte profiles in type 1 diabetics condition	Phase II	RPCT in adults with long-term T1D (mean 15 years) (NCT00607230)		C-peptide levels rose transiently in two BCG-treated subjects [115] Now endies (nJamed and recentifier)
Research trial of Aralast in new-onset diabetes (RETAIN-1)	Phases I/II	Open-label, safety and dose level study in adults and children with new onset T1D (NCT01183468)	http://www.immunetolerance.org/ studies/research-trial-aralast-	rew studies (painted and tecturing) Recruited, in follow-up
RETAIN-II Rituximab in early onset type 1 diabetes Vitamin D in the honeymoon period in children and adolescents with type 1 diabetes	Phase II Phase II	Planned RPCT of aralast in new onset 711D (NCT01183455) Open-label, Nanjing Medical University, China (NCT01280682) RPCT examining effect of vitamin D supplementation (3000 IU cholecalciferol daily for 9 months) on rate of partial clinical remission [assessed by insulin dose adjusted HbA1c	new-onsertuabetes-retain	In planning In recruitment In planning
Autologous mesenchymal stem cells in new onset	Phases I/II	Open-label safety and efficacy (NCT01068951)		In recruitment
Atorvastatin in New Onset Type 1 Diabetes Alefacept (soluble leucocyte function-associated antigen (EA)-3-1g fusion) in new-onset T1D	Phase II Phase II	RPCT (NCT00965458)		Ongoing, not recruiting Ongoing, not recruiting
Neulasta (GCSF) in type 1 diabetes Rilonacept (IL-1 trap) in diabetes mellitus type 1	Phases I/II Phase I	RPCT (NCT00662519) Open-label (NCT00962026)		Ongoing, not recruiting In recruitment
Immune therapy using CD4*CD127lo/-CD25*	Phase I	Open-label, dose escalation of Treg infusion (NCT01210664)		In recruitment
Autologous umbilical cord blood transfusion: pilot study	Phases I/II	EudraCT number: 2007-007694-23		In recruitment

RPCT: randomized placebo-controlled trial; DKA: diabetic ketoacidosis, FDA: US Food and Drug Administration; AHSCT: autologous haematopoietic stem cell transplantation; ATG: anti-thymocyte globulin; GCSF: granulocyte colony stimulating factor; IR: interleukin; mAb: monoclonal antibody; Treg: regulatory T cell.

Table 5. Completed, ongoing and planned prevention/intervention trials in type 1 diabetes (T1D) using combination approaches.

		,1, , , , , , , , , , , , , , , , , , ,		
Agent	Stage of development	Details (including ClinicalTrials.gov Identifier) Referen	References and links	
Low-dose cyclosporin and methotrexate Exenatide and Gastrin	Phases I/II Preclinical	Open-label; cyclosporin 7-5 mg/kg/day for 6 weeks and then 4 mg/kg/day Study comp for 1 year and methotrexate 5 mg/kg/day for 1 year (NCT00905073) no report http://www.transitiontherapeutics.com	Study completed; no report No further information	
			Progress to date	o date
Mycophenolate mofetil (MMF) and anti-CD25 mAb (Daclizumab; DZB)	Phase II	Multi-centre RPCT, new-onset T1D ( $n = 1267 < 3$ months) randomized to either MMF alone, MMF plus DZB or placebo. Mean C-peptide AUC at 2 years was unaffected by MMF alone or MMF plus DZB <i>versus</i> placebo (NCT00100178)	Study repo	Study reported [116]
Anti-CD3 and intranasal insulin	Phase II	Planned, recent-onset T1D	No further drug ac	No further progress; drug access difficulties
Anti-CD3 and Exenatide	Phase II	Various trials planned in recent onset T1D and in at-risk individuals (prevention)	No furthe drug ac	No further progress; drug access difficulties
Proleukin and Rapamune in type 1 diabetes	Phase I	Open-label; 9 recent-onset (6–48 months) T1D subjects treated with 2–4 mg/day rapamycin orally for 3 months and $4.5 \times 10^6$ IU IL-2 s.c. three times per week for 1 month showed transient treatment-induced $\beta$ cell dysfunction (NCT00525889	Study reported [20]	orted [20]
			New studies	es
Autologous stem cell transplantation for early-onset type 1 diabetes mellitus	Phases I/II	Open-label study of high-dose immunosuppression followed by autologous non-myeloablative haematopoietic stem cell transplantation (AHSCT) in newly diagnosed T1D (< 6 weeks; n = 15 patients aged 14–31 years) after conditioning with cyclophosphamide and rabbit ATG. During a 7–36-month follow-up (mean 18-8), 14 patients became insulin-free. At 6 months mean C-peptide response was significantly greater than the pretreatment values. Extension and follow-up shows a majority of patients became insulin-free and C-peptide levels increased significantly at 24 months. Two patients developed bilateral nosocomial pneumonia, 3 patients developed late endocrine dysfunction, and 9 patients developed oligospermia (NCT00315133)	[117–120]	
Haematopoietic stem cell transplantation in T1D	Phase II	Open label; AHSCT in new-onset T1D (NCT01121029)	Study repo	Study report awaited
Cord blood plus vitamin D and omega 3 s in T1D condition	Phase II	Open label randomized 2:1 (active: placebo); active is single intravenous infusion of autologous cord blood cells followed by 1 year of daily vitamin D and omega 3 fatty acid supplementation in 15 subjects; placebo untreated ( NCT00873925)	Study repo	Study report awaited
Efficacy and safety study of autologous haematopoietic stem cell transplantation to treat new onset type 1 diabetes	Phase I	Open label, non-myeloablative stem cell transplantation after conditioning with cyclophosphamide and rabbit ATG, Nanjing Medical University, China (NCT01341899)	In recruitment	nent
High-dose immunosuppression and AHSCT in early onset T1D	Phases I/II	Open label, < 5 months from diagnosis; protocol as for NCT00315133 to include rituximab conditioning and quality of life questionnaire (NCT01285934)	In recruitment	nent
Reversing type 1 diabetes after it is established	Phases I/II	Single-blind (participant) pilot safety and feasibility study of ATG and GCSF (Neulasta®) in established T1D (4–24 months) (NCT01106157)	Ongoing	

RCPT: randomized placebo-controlled trial; autologous haematopoietic stem cell transplantation; ATG: anti-thymocyte globulin; s.c.: subcutaneous; mAb: monoclonal antibody.

to certain immune intervention strategies [7]. While common beliefs advocate a combination of drugs for intervention (Table 5), it is important to scrutinize potential adverse interference, as may have played a role in the recent trial combining low-dose interleukin (IL)-2 and rapamycin, in which each of the separate constituents could have yielded clinical benefit [20]. Preclinical studies should be used carefully to identify those showing the desired synergy or any concerns in relation to the single components of combinations (i.e. accelerated disease, see below).

## Systemic immune modulators

Biological agents have proved to be immensely valuable in the treatment of autoimmune disease, and type 1 diabetes is no exception to this therapeutic track. Biologics targeting lymphocytes or co-stimulation events generally invoke immune suppression rather than modulation. This was perhaps most evident in case of the rituximab intervention study, in which patients were vaccinated under the treatment umbrella in a rare attempt to understand the mechanism of action of anti-CD20 immunotherapy. Indeed, rituximab blunted the induction of immune responses against a neoantigen, whereas after revaccination 1 year later (3 months after cessation of rituximab therapy) vigorous responses to the same neoantigen were established that did not differ from placebo-treated patients [21]. This observation underscores the fact that anti-CD20 therapy suppression possibly inhibits new immune responses but, as we know from the intervention study, does not instil or restore tolerance [8]. The term 'biologic cyclosporin' has been coined in this context. The recently reported failure of anti-thymocyte globulin to preserve C-peptide in a Phase II setting is a further wake-up call in this respect, emphasizing at the same time the complexity of human cellular autoimmune responsiveness and the bluntness of some of the tools at our disposal [22].

While biologics may prevent priming or spreading of the immune response, for most there is little evidence that they affect existing adaptive immunity. Indeed, abatacept [cytotoxic T lymphocyte antigen 4-immunoglobulin (CTLA4-Ig)] is effective at preventing priming alloreactivity, but appears to have little impact in reversing primed islet autoimmunity [14]. The reduced requirement for co-stimulation of autoreactive memory T cells [23] probably explains the limited clinical efficacy observed in the established disease process of chronic islet autoimmunity [14]. None the less, dimming immune reactivity with abatacept proved successful in delaying the progressive loss of stimulated C-peptide capacity in some patients in this study. The fact that the effect waned, even during continued treatment, again hints at disease heterogeneity, for example in the degree to which autoreactive T cell responses are co-stimulation-dependent.

With the exception of a small study using tumour necrosis factor (TNF)-α blockade [24], which showed potential clinical efficacy (which cannot currently be explored further due to safety concerns; see Table 4), interference in the activity of effector cytokines has not yet delivered in type 1 diabetes, as underlined by two recent failed studies of IL-1 blockade [25] (Table 4). This is in striking contrast with rheumatoid arthritis (e.g. benefits of blockade of TNF-α, IL-6 receptor, IL-1) and psoriasis (TNF-α, IL-23 and IL-17 pathways, IL-1). A central role for these cytokines in the immunopathogenesis may therefore be worthy of greater scrutiny and reconsideration, in spite of their clear role in some preclinical models of autoimmune diabetes and other autoimmune diseases. It remains plausible, of course, that cytokine inhibition will be highly effective and synergistic in combinations with other immune intervention strategies, as preclinical models imply [26].

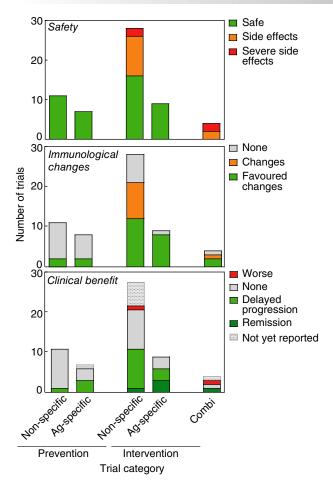
# Antigen-specific approaches

Viewed by many as the best chance to restore immunological self-tolerance in autoimmune diseases, antigen-specific immunotherapy (ASI) faces many challenges in its development and deployment, which is perhaps reflected in the more limited pipelines and activity in this arena (Tables 1 and 3; Fig. 1). Many of the relevant issues have been discussed elsewhere [27], but to put this modality into perspective several of the notable challenges are highlighted in Table 6. Perhaps in reflection of these, there has been limited new activity in this arena since 2007. Notably, a large primary prevention study of daily intranasal insulin reported failure to halt progression to type 1 diabetes [28], while the repeat oral insulin study conducted by Type 1 Diabetes TrialNet will not be able to report results until 2015, at the earliest. In the intervention setting, follow-up studies of alum-conjugated glutamic acid decarboxylase immunization (GAD-Alum), after initial successful pilot data [29], have been disappointing at Phase II [30] and Phase III stages [12]; a secondary prevention study is in progress (Table 1).

New modalities of ASI have emerged, however, including peptide and DNA-based deliveries, in some cases associated with positive biomarker data [16,31] and in the case of Diapep277, with evidence of clinical effectiveness (see discussion above and Table 3). Full reporting of the proinsulin-DNA vaccine and Diapep277 Phase III studies are eagerly awaited. In terms of development, however, it is notable that, for example, in the intervention setting, there has been no attempt as yet to combine antigen with any other treatment modality (Fig. 2), despite encouraging preclinical data [32,33].

# The role of preclinical models in trial design

With the somewhat high number of failed clinical trials in type 1 diabetes in the past few years, it has become increas-



**Fig. 1.** Bar charts show the number of clinical trials in different categories according to the natural history of type 1 diabetes (prevention, intervention) and treatment modality (none- or antigen-specific).

ingly tempting to attribute some of the blame to animal models. One often hears remarks such as 'animal models have misled us' and the near-ubiquitous comment 'mice are not humans'. Clearly, we are all aware that diabetes in various rodent models may only model in part how type 1 diabetes develops in humans. However, we would like to argue here that animal models have a key place in the clinical translation for therapeutic approaches in autoimmune disease overall, as long as they are used correctly, not overinterpreted and analysed carefully. It should be helpful, therefore, to first take a closer look at the extent to which animal studies diverge from human trials.

Several ASI trials in man have reported negative (or positive substudy) results (GAD-Alum, oral insulin and intravenous insulin); have shown marginal effects (BayHill DNA vaccine, Diapep277); or were not powered to demonstrate efficacy, yet have not shown any strong clinical effects in established diabetes (adjuvanted insulin B-chain peptide, proinsulin peptide). Each trial is distinctly different and it is therefore worthwhile to look at the facts one by one.

Subcutaneous administration of GAD-Alum was developed on the basis of earlier studies by several teams, which had all used GAD peptides to prevent diabetes in the nonobese diabetic (NOD) mouse spontaneous disease model [34,35]. Others have since prevented type 1 diabetes successfully with oral GAD and in some cases GAD DNA vaccines also using other diabetes models [36]. A crucial difference between the human trial and all the preclinical studies is that immunization with GAD always worked to prevent diabetes, yet never after diabetes onset. As discussed, this is a universal truth for all ASI, which has not shown disease-reverting effects in animal models after clinical signs of diabetes have developed. Thus, it would be unreasonable to expect a stronger effect (in other words, after onset of diabetes) in humans. Secondly, no preclinical study ever tested the clinical GAD-Alum preparation, and no efficacy was noted in our recent studies in NOD and B6 diabetes models (Pagni, Boettler and von Herrath, unpublished). Again, it is probably unreasonable to expect an antigenic formulation to work in humans when it does not even prevent diabetes in otherwise permissive animal models. Several other theories have been proposed to account for the failure of GAD-Alum in humans, including the lack of GAD expression in  $\beta$  cells; this is a controversial area, as many studies have demonstrated expression of GAD-65 and 67 proteins in murine and human  $\beta$  cells [36]. Lastly, one could ask whether the dose of GAD-Alum was sufficient as most patients mounted a clearly detectable immune response, this appears less likely. However, alum might have been a suboptimal adjuvant for an ASI, as the resulting mixed but T helper type 2 (Th2)-dominated cytokine response of induced GAD-reactive T cells (Arif, Roep and Peakman, unpublished) did not result in protective cell populations. In the absence of a functional mouse model of GAD-Alum preventing diabetes, it will be difficult at this point to clarify these issues.

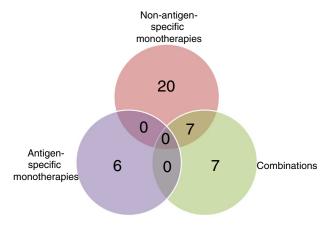
The question of the antigenic dose might have more bearing on the issue of efficacy with oral insulin [15]. As predicted from animal models [37], prophylactic oral insulin given at a daily dose of 7.5 mg had a very marginal effect in preventing diabetes in individuals at high risk (exhibiting multiple autoantibodies [38-41]), but not in any other patient groups. However, as has been evident from multiple studies in different mouse models, oral insulin dosages have to be comparatively much higher to induce optimal disease preventive effects, which are seen at a dose of 1 mg given twice per week [42]. This dose would equate to approximately 1 g of oral insulin twice per week in humans. In addition, it is likely to be necessary to provide the drug in enteric-coated capsules, without which > 99.99% of the insulin is lost through digestion in the stomach and only minimal amounts of intact antigen or some peptides will reach the lower gut and the Peyer's patches, the location at which oral insulin has been shown to induce its desired immune-regulatory response. There-

Table 6. Challenges faced in the development of antigen-specific immunotherapy (ASI) for type 1 diabetes.

Challenge	Discussion of issues
Setting for clinical trials	Traditionally, new therapies are trialled in the intervention setting (i.e. at disease onset). Disease reversal using antigen alone at this stage will most probably be difficult. Prevention studies are long duration and expensive; but without hints of efficacy as an intervention, will prevention studies be undertaken?
Dose	Both high- and low-dose immunological tolerance has been described, probably equating to predominantly deletional and regulatory mechanisms; which is better, and whether both effects could be harnessed, is not known, however
Regime	Frequent (daily) dosing has been the norm until now (e.g. for intranasal and oral insulin), but again this may favour deletion over regulation [27]
Adjuvants and enhancing combinations	A poorly explored area in general, despite encouraging data in preclinical models (e.g. anti-CD3 plus antigen; see Table 5)
Agent	It has yet to be determined whether whole antigens or fragments are superior; similarly, whether protein or DNA-based delivery is better; free peptide or complexed to peptide–human leucocyte antigen multimers or nanoparticles
Route of administration	Parenteral or oral/nasal routes predominate, but the relative advantages of either have not been explored head-to-head
Staging and stratification	Oral insulin appears effective in the subgroup of patients with high titres of insulin autoantibodies; is this a general principle for ASI?
Preclinical models	As a generalization, ASI works well if given early enough in disease models; but trialling the human antigens in humanized models is an under-developed area
Role of industry and biotech	Antigens face the dual challenges of being difficult to develop with robust intellectual property and having a clear route to market and have therefore been less favoured for commercial development than biologics and other immune modulators

fore, more precise dose calculations should have probably preceded the oral insulin trial and its current follow-up study.

A further human/mouse mismatch relates to the overall management of expectations when devising trials for ASI. In rodent studies most, if not all, ASI is effective only for early and, at best, late prevention of disease, but never after onset of hyperglycaemia. Thus, we should not expect antigens to reverse human diabetes or even preserve C-peptide after onset (at least with effects detectable in reasonably sized studies); and this has indeed been the case. Rather, these types of studies are potentially invaluable for optimizing dose and administration schemes and biomarker devel-



**Fig. 2.** Venn diagram shows the number of types of therapies used in different modalities (mono- and combination; antigen and non-antigen-specific) and the overlap between them.

opment, if immunological parameters are used as an outcome. Here there will need to be 'reverse translation', because immune parameters are analysed rarely on peripheral blood and correlated with successful prevention (or lack thereof) of diabetes on an individual basis in murine studies.

Surprisingly, two recent trials (Andromeda's heat shock protein peptide p277 and Bayhill's proinsulin expressing DNA vaccine BHT3021; Table 4) reported positive outcomes, even in the more stringent recent-onset diabetes setting, by preserving C-peptide at certain dosing regimens. These observations exceeded expectations based on animal studies, where both strategies were only effective in preventing diabetes but not in reversing hyperglycaemia. It will be important to explore whether, in either trial, immunological outcomes were associated with better preservation of C-peptide and thus could perhaps pave the way in future for using such immunological end-points in staging as entry criteria, or to optimize dosing in larger trials, prior to embarking on the more arduous, expensive and time-consuming prevention trials.

# What we do better now or know better now?

Recent, seminal lessons from studies on pancreatic tissue of type 1 diabetic donors provide compelling proof of the autoimmune nature of type 1 diabetes; in particular, the demonstration of  $\beta$  cell autoantigen-specific CD8 T cells in destructive insulitic lesions has highlighted a link that had not emerged in 2007. The persistence of  $\beta$  cells and insulin production as well as inflammatory insulitic lesions many

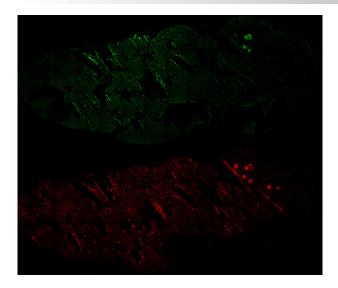


Fig. 3. Frozen pancreas tissue section from an organ donor who was diagnosed with type 1 (T1D) diabetes 1 year earlier. The section was stained for insulin (green, top) and human leucocyte antigen (HLA)-ABC (red, bottom). Individual images were captured by confocal microscopy and automatically combined *in-silico* into a section-wide overview figure. Note the distinct region in the upper right corner where some insulin-producing beta cells are still present. These remaining insulin-positive islets also hyperexpress major histocompatibility complex (MHC) class I, as evident in the lower panel. Thus, this is a prime example of the lobular distribution of both beta cell loss and immune pathology in T1D. Samples courtesy of Network for Pancreatic Organ Donors with Diabetes (nPOD) (case 6052) and kindly provided by Dr Ken Coppieters.

years after clinical manifestations of hyperglycaemia are also arresting, providing an apparent disconnect between  $\beta$  cell mass and function. These studies also emphasize differences in immunopathology between men and mice; provide evidence of pathological and aetiological heterogeneity [43–49]; and provide potential new biomarkers and therapeutic targets centred on CD8 T cell biology [50–53] that were not envisaged at the time of our last review-(Fig. 3). Importantly, the 'biomarker concept' that has become a critical piece of new drug development in the pharma industry has also begun to feature strongly in current thinking about type 1 diabetes therapies [5]; the term was not even used in the previous paper [1].

There is probably more new insight to be gained from studying the diabetic pancreas in settings such as nPOD. For example, the observation that the remaining  $\beta$  cell mass at clinical manifestation of disease may be substantial (as much as 50%, rather than 10–20% cited in most textbooks) disproves a common assumption that the disease process has always reached an end-stage at this point.

Immune monitoring in clinical islet transplantation has shown that the current immune suppressive regimes that include thymoglobulin are ineffective for the control of memory T cells; this concept is supported further by the lack of efficacy observed in the recent anti-thymocyte globulin (ATG) trial (Table 4) [54]. Thus the autoreactive memory T cell, and the nature of its biology and control, emerge as important research questions, built on knowledge gained in recent years. As discussed already, the disappointing outcome of trials targeting the proinflammatory cytokine IL-1 [25] may require a revision of thinking in relation to the importance of this immune pathway. Finally, a relatively new paradigm has come to prominence, namely that the biology of  $\beta$  cells can contribute to the cell's own demise through active participation at key points of the interface with the immune system, from immune recognition to immune cell recruitment and killing [55-57]. A better understanding of these processes could be useful in devising better combination-based candidate strategies of immune intervention and prevention in type 1 diabetes.

# The future pipeline

We would like to argue that animal models, when employed correctly, can be extremely useful for testing and optimizing new interventions for human type 1 diabetes. In addition, the new knowledge being accrued must be assimilated. We suggest the following strategic guidelines for pipeline development.

- 1. Defining the optimal dose for an antigen or biologic. Treating with the correct dose is of paramount importance, for ASI treatment with incorrect doses may result in loss of efficacy (see above) or may even be accelerating. For biologics, treating at an incorrect dose may not only mean loss of effect (as with otelixizumab in Phase III), but also increased side effects, if too much drug is given. Assumptions may be made that, for example, a monoclonal antibody targeting T cells will be effective as long as there is target molecule internalization; however, studies in mice show that there may be an approximate log-fold difference in dose between internalization and full efficacy. Thus, careful dosing studies in models, coupled with appropriate biomarkers, will be critical in attaining good efficacy in humans.
- 2. Preclinical testing of combinations. Despite the logic of this approach, it is becoming clear that not all combinations exhibit additive effects, let alone synergies. Thus, careful optimization of combinations prior to clinical trials is needed. As a case in point, for example, not all antigens synergize with anti-CD3 therapy [32]. To accelerate translation in this arena, the Immune Tolerance Network (ITN; http://www.immunetolerance.org) has established a combination therapy testing consortium, in which four independent laboratories evaluate combinations of biologics and antigens in recent-onset diabetes in NOD mice. Such studies have so far demonstrated limited additive effects when examining potentially new combinations of biologics and antigens in recent-onset

- diabetes. Clearly, it will be important to establish which combinations work, and how.
- 3. Assessing patient heterogeneity. Is all type 1 diabetes the same? Our knowledge to date indicates that this is unlikely to be the case, and this should caution us to anticipate subgroup effects. For example, the rate of  $\beta$ cell loss varies between individuals, being most rapid in younger individuals aged 20 or less [58]. The fact that Diapep277 only had its effects in older patients and in those with lower-risk major histocompatibility complex (MHC) illustrates this [59]. To date, we are not certain whether the underlying immune pathology varies between different forms of type 1 diabetes (for example, a more IFN-dependent versus a more IL-dependent diabetes, or T cell-dependent versus NK cell-dependent islet destruction [60]), but nPOD studies may elucidate this. In that case, stratifying patients by immune phenotypes may become an increasingly important feature of trial design.
- 4. Defining the optimal disease stage for a given therapy. One paradigm that may emerge from ongoing diabetes trials is that the more aggressive the immune CD8 reactivity to islets, the more advanced  $\beta$  cell loss is, the less likely it is that any treatment will be effective (various studies, unpublished). Monoclonal anti-CD3 antibodies do not appear to preserve C-peptide in patients with advanced  $\beta$  cell loss (lower C-peptide at trial entry).
- 5. Managing expectations. Taking the above issues at face value, not over-interpreting the data from animal models or being excessively optimistic ('this has to work') and refraining from conducting trials simply because drugs are available and effective in other immune disorders is an important message set to help avoidance of disappointments with future diabetes trials.

# Acknowledgements

M.P. acknowledges support from the UK Department of Health via the National Institute for Health Research (NIHR) Biomedical Research Centre award to Guy's & St Thomas' NHS Foundation Trust in partnership with King's College London. M.P. and B.O.R. receive funding via the EU FP7 Framework 7 Large-scale Focused Collaborative Research Project on Natural Immunomodulators as Novel Immunotherapies for Type 1 diabetes (NAIMIT) and EE-ASI (Beta cell preservation via antigen-specific immunotherapy in Type 1 Diabetes: Enhanced Epidermal Antigen Delivery Systems); M.P. is also funded via the EU FP7 PEVNET programme (Persistent virus infection as a cause of pathogenic inflammation in type 1 diabetes - an innovative research program of biobanks and expertise) and as part of the Juvenile Diabetes Research Foundation Autiommunity Centers Consortium (1-2007-1803). We are grateful to Dr Ken Coppieters for providing the image used in Fig. 3. This image was generated with the support of the Network for Pancreatic Organ Donors with Diabetes (nPOD), a collaborative type 1 diabetes research project sponsored by the Juvenile Diabetes Research Foundation International. Organ Procurement Organizations (OPO) partnering with nPOD to provide research resources are listed at http://www.jdrfnpod.org/our-partners.php.

#### References

- 1 Staeva-Vieira T, Peakman M, von Herrath M. Translational minireview series on type 1 diabetes: immune-based therapeutic approaches for type 1 diabetes. Clin Exp Immunol 2007; 148:17– 31
- 2 Herold KC, Hagopian W, Auger JA et al. Anti-CD3 monoclonal antibody in new-onset type 1 diabetes mellitus. N Engl J Med 2002; 346:1692–8.
- 3 Keymeulen B, Vandemeulebroucke E, Ziegler AG *et al.* Insulin needs after CD3-antibody therapy in new-onset type 1 diabetes. N Engl J Med 2005; **352**:2598–608.
- 4 Matthews JB, Staeva TP, Bernstein PL, Peakman M, von Herrath M. Developing combination immunotherapies for type 1 diabetes: recommendations from the ITN–JDRF Type 1 Diabetes Combination Therapy Assessment Group. Clin Exp Immunol 2010; 160:176–84.
- 5 Roep BO, Peakman M. Surrogate end points in the design of immunotherapy trials: emerging lessons from type 1 diabetes. Nat Rev Immunol 2010; 10:145–52.
- 6 Coppieters KT, Dotta F, Amirian N et al. Demonstration of isletautoreactive CD8 T cells in insulitic lesions from recent onset and long-term type 1 diabetes patients. J Exp Med 2012; 209:51–60.
- 7 Ahmed S, Akirav E, Bradshaw EM et al. Immunologic biomarkers: catalysts for translational advances in autoimmune diabetes. Clin Exp Immunol 2013; 172:178–85.
- 8 Pescovitz MD, Greenbaum CJ, Krause-Steinrauf H et al. Rituximab, B-lymphocyte depletion, and preservation of beta-cell function. N Engl J Med 2009; 361:2143–52.
- 9 Keymeulen B, Walter M, Mathieu C et al. Four-year metabolic outcome of a randomised controlled CD3-antibody trial in recent-onset type 1 diabetic patients depends on their age and baseline residual beta cell mass. Diabetologia 2010; 53:614–23.
- 10 Campbell-Thompson M, Wasserfall C, Kaddis J et al. Network for Pancreatic Organ Donors with Diabetes (nPOD): developing a tissue biobank for type 1 diabetes. Diabetes Metab Res Rev 2012; 28:608–17.
- 11 Meier JJ, Bhushan A, Butler AE, Rizza RA, Butler PC. Sustained beta cell apoptosis in patients with long-standing type 1 diabetes: indirect evidence for islet regeneration? Diabetologia 2005; 48:2221–8.
- 12 Ludvigsson J, Krisky D, Casas R et al. GAD65 antigen therapy in recently diagnosed type 1 diabetes mellitus. N Engl J Med 2012; 366:433–42.
- 13 Sherry N, Hagopian W, Ludvigsson J *et al*. Teplizumab for treatment of type 1 diabetes (Protege study): 1-year results from a randomised, placebo-controlled trial. Lancet 2011; **378**:487–97.
- 14 Orban T, Bundy B, Becker DJ et al. Co-stimulation modulation with abatacept in patients with recent-onset type 1 diabetes: a randomised, double-blind, placebo-controlled trial. Lancet 2011; 378:412–9.

- 15 Skyler JS, Krischer JP, Wolfsdorf J *et al.* Effects of oral insulin in relatives of patients with type 1 diabetes: the Diabetes Prevention Trial Type 1. Diabetes Care 2005; **28**:1068–76.
- 16 Thrower SL, James L, Hall W *et al.* Proinsulin peptide immunotherapy in type 1 diabetes: report of a first-in-man Phase I safety study. Clin Exp Immunol 2009; **155**:156–65.
- 17 Huurman VA, van der Torren CR, Gillard P *et al*. Immune responses against islet allografts during tapering of immunosuppression a pilot study in five subjects. Clin Exp Immunol 2011; **169**:190–8.
- 18 Hilbrands R, Huurman VA, Gillard P *et al.* Differences in baseline lymphocyte counts and autoreactivity are associated with differences in outcome of islet cell transplantation in type 1 diabetic patients. Diabetes 2009; **58**:2267–76.
- 19 Huurman VA, Hilbrands R, Pinkse GG et al. Cellular islet autoimmunity associates with clinical outcome of islet cell transplantation. PLoS ONE 2008; 3:e2435.
- 20 Long SA, Rieck M, Sanda S *et al.*, Diabetes TrialNet and the Immune Tolerance Network. Rapamycin/IL-2 combination therapy in patients with type 1 diabetes augments Tregs yet transiently impairs beta-cell function. Diabetes 2012; **61**:2340–8.
- 21 Pescovitz MD, Torgerson TR, Ochs HD et al. Type 1 Diabetes TrialNet Study G. Effect of rituximab on human in vivo antibody immune responses. J Allergy Clin Immunol 2011; 128:1295–302.
- 22 Gitelman SE, Fisher LK, Gottlieb PA et al., START Study Team. Effect of anti-thymocyte globulin (ATG) on preserving beta cell function in new-onset type 1 diabetes. Poster Presentation 454, European Association for the Study of Diabetes, 2012.
- 23 Viglietta V, Kent SC, Orban T, Hafler DA. GAD65-reactive T cells are activated in patients with autoimmune type 1a diabetes. J Clin Invest 2002; 109:895–903.
- 24 Mastrandrea L, Yu J, Behrens T et al. Etanercept treatment in children with new-onset type 1 diabetes: pilot randomized, placebo-controlled, double-blind study. Diabetes Care 2009; 32:1244–9.
- 25 Moran A, Bundy B, Becker DJ *et al.*, Group AS. Interleukin-1 antagonism in type 1 diabetes of recent onset: two multicenter, randomized double-masked, placebo-controlled trials. Lancet 2013; in press.
- 26 Ablamunits V, Henegariu O, Hansen JB *et al.* Synergistic reversal of type 1 diabetes in NOD mice with anti-CD3 and interleukin-1 blockade: evidence of improved immune regulation. Diabetes 2012: **61**:145–54
- 27 Tree TI, Lawson J, Edwards H et al. Naturally arising human CD4 T-cells that recognize islet autoantigens and secrete interleukin-10 regulate proinflammatory T-cell responses via linked suppression. Diabetes 2010; 59:1451–60.
- 28 Nanto-Salonen K, Kupila A, Simell S et al. Nasal insulin to prevent type 1 diabetes in children with HLA genotypes and autoantibodies conferring increased risk of disease: a double-blind, randomised controlled trial. Lancet 2008; 372:1746–55.
- 29 Ludvigsson J, Faresjo M, Hjorth M et al. GAD treatment and insulin secretion in recent-onset type 1 diabetes. N Engl J Med 2008: 359:1909–20.
- 30 Wherrett DK, Bundy B, Becker DJ *et al.* Antigen-based therapy with glutamic acid decarboxylase (GAD) vaccine in patients with recent-onset type 1 diabetes: a randomised double-blind trial. Lancet 2011; **378**:319–27.
- 31 Orban T, Farkas K, Jalahej H et al. Autoantigen-specific regulatory T cells induced in patients with type 1 diabetes mellitus

- by insulin B-chain immunotherapy. J Autoimmun 2010; **34**: 408–15.
- 32 Bresson D, Togher L, Rodrigo E et al. Anti-CD3 and nasal proinsulin combination therapy enhances remission from recent-onset autoimmune diabetes by inducing Tregs. J Clin Invest 2006; 116:1371–81.
- 33 Takiishi T, Korf H, Van Belle TL et al. Reversal of autoimmune diabetes by restoration of antigen-specific tolerance using genetically modified *Lactococcus lactis* in mice. J Clin Invest 2012; 122:1717–25.
- 34 Kaufman DL, Clare-Salzler M, Tian J et al. Spontaneous loss of T-cell tolerance to glutamic acid decarboxylase in murine insulindependent diabetes. Nature 1993; 366:69–72.
- 35 Tisch R, Yang XD, Singer SM, Liblau RS, Fugger L, McDevitt HO. Immune response to glutamic acid decarboxylase correlates with insulitis in non-obese diabetic mice. Nature 1993; 366:72–5.
- 36 Bresson D, Fradkin M, Manenkova Y, Rottembourg D, von Herrath M. Genetic-induced variations in the GAD65 T-cell repertoire governs efficacy of anti-CD3/GAD65 combination therapy in new-onset type 1 diabetes. Mol Ther 2010; 18:307–16.
- 37 Fousteri G, Chan JR, Zheng Y *et al.* Virtual optimization of nasal insulin therapy predicts immunization frequency to be crucial for diabetes protection. Diabetes 2010; **59**:3148–58.
- 38 Gorus FK, Balti EV, Vermeulen I *et al.* Screening for insulinoma antigen 2 and zinc transporter 8 autoantibodies: a cost-effective and age-independent strategy to identify rapid progressors to clinical onset among relatives of type 1 diabetic patients. Clin Exp Immunol 2013; 171:82–90.
- 39 Gabbay MA, Sato MN, Duarte AJ, Dib SA. Serum titres of anti-glutamic acid decarboxylase-65 and anti-IA-2 autoantibodies are associated with different immunoregulatory milieu in newly diagnosed type 1 diabetes patients. Clin Exp Immunol 2012; 168:60–7.
- 40 Curnock RM, Reed CR, Rokni S, Broadhurst JW, Bingley PJ, Williams AJ. Insulin autoantibody affinity measurement using a single concentration of unlabelled insulin competitor discriminates risk in relatives of patients with type 1 diabetes. Clin Exp Immunol 2012; **167**:67–72.
- 41 Tiberti C, Yu L, Lucantoni F et al. Detection of four diabetes specific autoantibodies in a single radioimmunoassay: an innovative high-throughput approach for autoimmune diabetes screening. Clin Exp Immunol 2011; 166:317–24.
- 42 Peakman M, von Herrath M. Antigen-specific immunotherapy for type 1 diabetes: maximizing the potential. Diabetes 2010; 59:2087–93.
- 43 Hober D, Sane F, Jaidane H, Riedweg K, Goffard A, Desailloud R. Immunology in the Clinic Review Series; focus on type 1 diabetes and viruses: role of antibodies enhancing the infection with Coxsackievirus-B in the pathogenesis of type 1 diabetes. Clin Exp Immunol 2012; 168:47–51.
- 44 Jaidane H, Sane F, Hiar R *et al.* Immunology in the Clinic Review Series; focus on type 1 diabetes and viruses: enterovirus, thymus and type 1 diabetes pathogenesis. Clin Exp Immunol 2012; **168**:39–46.
- 45 Lind K, Huhn MH, Flodstrom-Tullberg M. Immunology in the Clinic Review Series; focus on type 1 diabetes and viruses: the innate immune response to enteroviruses and its possible role in regulating type 1 diabetes. Clin Exp Immunol 2012; 168:30–8.
- 46 Grieco FA, Sebastiani G, Spagnuolo I, Patti A, Dotta F. Immunology in the Clinic Review Series; focus on type 1 diabetes and

- viruses: how viral infections modulate beta cell function. Clin Exp Immunol 2012; **168**:24–9.
- 47 Stene LC, Rewers M. Immunology in the Clinic Review Series; focus on type 1 diabetes and viruses: the enterovirus link to type 1 diabetes: critical review of human studies. Clin Exp Immunol 2012; 168:12–23.
- 48 Coppieters KT, Wiberg A, Tracy SM, von Herrath MG. Immunology in the Clinic Review Series: focus on type 1 diabetes and viruses: the role of viruses in type 1 diabetes: a difficult dilemma. Clin Exp Immunol 2012; **168**:5–11.
- 49 Gale EA. Viruses and type 1 diabetes: ignorance acquires a better vocabulary. Clin Exp Immunol 2012; **168**:1–4.
- 50 Han D, Cai X, Wen J *et al.* Innate and adaptive immune gene expression profiles as biomarkers in human type 1 diabetes. Clin Exp Immunol 2012; **170**:131–8.
- 51 Abreu JR, Martina S, Verrijn Stuart AA et al. CD8 T cell autoreactivity to preproinsulin epitopes with very low human leucocyte antigen class I binding affinity. Clin Exp Immunol 2012; 170:57–65.
- 52 Coppieters KT, Amirian N, von Herrath MG. Incidental CD8 T cell reactivity against caspase-cleaved apoptotic self-antigens from ubiquitously expressed proteins in islets from prediabetic human leucocyte antigen-A2 transgenic non-obese diabetic mice. Clin Exp Immunol 2011; **165**:155–62.
- 53 Mallone R, Mannering SI, Brooks-Worrell BM *et al.* Isolation and preservation of peripheral blood mononuclear cells for analysis of islet antigen-reactive T cell responses: position statement of the T-Cell Workshop Committee of the Immunology of Diabetes Society. Clin Exp Immunol 2011; **163**:33–49.
- 54 Bresson D, von Herrath MG. Anti-thymoglobulin (ATG) treatment does not reverse type 1 diabetes in the acute virally induced rat insulin promoter–lymphocytic choriomeningitis virus (RIP–LCMV) model. Clin Exp Immunol 2011; 163:375–80.
- 55 Skowera A, Ellis RJ, Varela-Calvino R *et al.* CTLs are targeted to kill beta cells in patients with type 1 diabetes through recognition of a glucose-regulated preproinsulin epitope. J Clin Invest 2008; 118:3390–402.
- 56 Knight RR, Kronenberg D, Zhao M et al. Human beta-cell killing by autoreactive preproinsulin-specific CD8 T cells is predominantly granule-mediated with the potency dependent upon T-cell receptor avidity. Diabetes 2013; 62:205–13.
- 57 Kronenberg D, Knight RR, Estorninho M *et al.* Circulating preproinsulin signal peptide-specific CD8 T cells restricted by the susceptibility molecule HLA-A24 are expanded at onset of type 1 diabetes and kill beta-cells. Diabetes 2012; **61**:1752–9.
- 58 Greenbaum CJ, Beam CA, Boulware D et al. Fall in C-peptide during first 2 years from diagnosis: evidence of at least two distinct phases from composite Type 1 Diabetes TrialNet data. Diabetes 2012; 61:2066–73.
- 59 Buzzetti R, Cernea S, Petrone A et al. C-peptide response and HLA genotypes in subjects with recent-onset type 1 diabetes after immunotherapy with DiaPep277: an exploratory study. Diabetes 2011; 60:3067–72.
- 60 Simoni Y, Diana J, Ghazarian L, Beaudoin L, Lehuen A. Therapeutic manipulation of natural killer (NK) T cells in autoimmunity: are we close to reality? Clin Exp Immunol 2013; 171:8–19.
- 61 Keller RJ, Eisenbarth GS, Jackson RA. Insulin prophylaxis in individuals at high risk of type I diabetes. Lancet 1993; 341:927–8.
- 62 Fuchtenbusch M, Rabl W, Grassl B, Bachmann W, Standl E, Ziegler AG. Delay of type I diabetes in high risk, first degree

- relatives by parenteral antigen administration: the Schwabing Insulin Prophylaxis Pilot Trial. Diabetologia 1998; **41**:536–41.
- 63 Diabetes Prevention Trial-Type 1 Diabetes Study Group. Effects of insulin in relatives of patients with type 1 diabetes mellitus. N Engl J Med 2002; 346:1685–91.
- 64 Harrison LC, Honeyman MC, Steele CE *et al.* Pancreatic beta-cell function and immune responses to insulin after administration of intranasal insulin to humans at risk for type 1 diabetes. Diabetes Care 2004; **27**:2348–55.
- 65 Kupila A, Sipila J, Keskinen P *et al.* Intranasally administered insulin intended for prevention of type 1 diabetes a safety study in healthy adults. Diabetes Metab Res Rev 2003; **19**:415–20.
- 66 Fourlanos S, Perry C, Gellert SA et al. Evidence that nasal insulin induces immune tolerance to insulin in adults with autoimmune diabetes. Diabetes 2011; 60:1237–45.
- 67 Achenbach P, Barker J, Bonifacio E. Modulating the natural history of type 1 diabetes in children at high genetic risk by mucosal insulin immunization. Curr Diab Rep 2008; 8:87–93.
- 68 Bohmer KP, Kolb H, Kuglin B et al. Linear loss of insulin secretory capacity during the last six months preceding IDDM. No effect of antiedematous therapy with ketotifen. Diabetes Care 1994; 17:138–41.
- 69 Carel JC, Boitard C, Eisenbarth G, Bach JF, Bougneres PF. Cyclosporine delays but does not prevent clinical onset in glucose intolerant pre-type 1 diabetic children. J Autoimmun 1996; 9:739– 45.
- 70 Lampeter EF, Klinghammer A, Scherbaum WA et al. The Deutsche Nicotinamide Intervention Study: an attempt to prevent type 1 diabetes. DENIS Group. Diabetes 1998; 47:980–4.
- 71 Gale EA, Bingley PJ, Emmett CL, Collier T. European Nicotinamide Diabetes Intervention Trial (ENDIT): a randomised controlled trial of intervention before the onset of type 1 diabetes. Lancet 2004; 363:925–31.
- 72 Crino A, Schiaffini R, Ciampalini P et al. A two year observational study of nicotinamide and intensive insulin therapy in patients with recent onset type 1 diabetes mellitus. J Pediatr Endocrinol Metab 2005; 18:749–54.
- 73 Crino A, Schiaffini R, Manfrini S et al. A randomized trial of nicotinamide and vitamin E in children with recent onset type 1 diabetes (IMDIAB IX). Eur J Endocrinol 2004; 150: 719–24.
- 74 Pozzilli P, Visalli N, Boccuni ML et al. Randomized trial comparing nicotinamide and nicotinamide plus cyclosporin in recent onset insulin-dependent diabetes (IMDIAB 1). The IMDIAB Study Group. Diabet Med 1994; 11:98–104.
- 75 Dahlquist G, Gothefors L. The cumulative incidence of childhood diabetes mellitus in Sweden unaffected by BCG-vaccination. Diabetologia 1995; 38:873–4.
- 76 Huppmann M, Baumgarten A, Ziegler AG, Bonifacio E. Neonatal bacille Calmette–Guérin vaccination and type 1 diabetes. Diabetes Care 2005; 28:1204–6.
- 77 Parent ME, Siemiatycki J, Menzies R, Fritschi L, Colle E. Bacille Calmette–Guérin vaccination and incidence of IDDM in Montreal, Canada. Diabetes Care 1997; 20:767–72.
- 78 Hummel M, Bonifacio E, Naserke HE, Ziegler AG. Elimination of dietary gluten does not reduce titers of type 1 diabetes-associated autoantibodies in high-risk subjects. Diabetes Care 2002; 25:1111–6.
- 79 Wicklow BA, Taback SP. Feasibility of a type 1 diabetes primary prevention trial using 2000 IU vitamin D3 in infants from the

- general population with increased HLA-associated risk. Ann NY Acad Sci 2006; 1079:310–2.
- 80 Akerblom HK, Virtanen SM, Ilonen J *et al.* Dietary manipulation of beta cell autoimmunity in infants at increased risk of type 1 diabetes: a pilot study. Diabetologia 2005; **48**:829–37.
- 81 Knip M, Virtanen SM, Seppa K et al. Dietary intervention in infancy and later signs of beta-cell autoimmunity. N Engl J Med 2010; 363:1900–8.
- 82 Miller MR, Seifert J, Szabo NJ, Clare-Salzler M, Rewers M, Norris JM. Erythrocyte membrane fatty acid content in infants consuming formulas supplemented with docosahexaenoic acid (DHA) and arachidonic acid (ARA): an observational study. Matern Child Nutr 2010; 6:338–46.
- 83 Miller MR, Yin X, Seifert J *et al.* Erythrocyte membrane omega-3 fatty acid levels and omega-3 fatty acid intake are not associated with conversion to type 1 diabetes in children with islet autoimmunity: the Diabetes Autoimmunity Study in the Young (DAISY). Pediatr Diabetes 2011; **12**:669–75.
- 84 Vaarala O, Ilonen J, Ruohtula T *et al.* Removal of bovine insulin from cow's milk formula and early initiation of beta-cell autoimmunity in the FINDIA pilot study. Arch Pediatr Adolesc Med 2012; **166**:608–14.
- 85 Hummel S, Pfluger M, Hummel M, Bonifacio E, Ziegler AG. Primary dietary intervention study to reduce the risk of islet autoimmunity in children at increased risk for type 1 diabetes: the BABYDIET study. Diabetes Care 2011; **34**:1301–5.
- 86 Alleva DG, Maki RA, Putnam AL *et al.* Immunomodulation in type 1 diabetes by NBI-6024, an altered peptide ligand of the insulin B epitope. Scand J Immunol 2006; **63**:59–69.
- 87 Raz I, Elias D, Avron A, Tamir M, Metzger M, Cohen IR. Beta-cell function in new-onset type 1 diabetes and immunomodulation with a heat-shock protein peptide (DiaPep277): a randomised, double-blind, Phase II trial. Lancet 2001; 358: 1749–53.
- 88 Lazar L, Ofan R, Weintrob N et al. Heat-shock protein peptide DiaPep277 treatment in children with newly diagnosed type 1 diabetes: a randomised, double-blind Phase II study. Diabetes Metab Res Rev 2006; 23:286–91.
- 89 Agardh CD, Cilio CM, Lethagen A et al. Clinical evidence for the safety of GAD65 immunomodulation in adult-onset autoimmune diabetes. J Diabetes Complications 2005; 19:238–46.
- 90 Peakman M, Stevens EJ, Lohmann T et al. Naturally processed and presented epitopes of the islet cell autoantigen IA-2 eluted from HLA-DR4. J Clin Invest 1999; 104:1449–57.
- 91 Bougneres PF, Landais P, Boisson C *et al.* Limited duration of remission of insulin dependency in children with recent overt type I diabetes treated with low-dose cyclosporin. Diabetes 1990; **39**:1264–72.
- 92 Jenner M, Bradish G, Stiller C, Atkison P. Cyclosporin A treatment of young children with newly-diagnosed type 1 (insulindependent) diabetes mellitus. London Diabetes Study Group. Diabetologia 1992; 35:884–8.
- 93 Elliott RB, Chase HP. Prevention or delay of type 1 (insulindependent) diabetes mellitus in children using nicotinamide. Diabetologia 1991; **34**:362–5.
- 94 Eisenbarth GS, Srikanta S, Jackson R *et al.* Anti-thymocyte globulin and prednisone immunotherapy of recent onset type 1 diabetes mellitus. Diabetes Res 1985; 2:271–6.
- 95 Allen HF, Klingensmith GJ, Jensen P, Simoes E, Hayward A, Chase HP. Effect of bacillus Calmette–Guérin vaccination on new-onset

- type 1 diabetes. A randomized clinical study. Diabetes Care 1999; **22**:1703–7.
- 96 Elliott JF, Marlin KL, Couch RM. Effect of bacille Calmette–Guérin vaccination on C-peptide secretion in children newly diagnosed with IDDM. Diabetes Care 1998; 21:1691–3.
- 97 Shehadeh N, Calcinaro F, Bradley BJ, Bruchim I, Vardi P, Lafferty KJ. Effect of adjuvant therapy on development of diabetes in mouse and man. Lancet 1994; 343:706–7.
- 98 Bjork E, Berne C, Kampe O, Wibell L, Oskarsson P, Karlsson FA. Diazoxide treatment at onset preserves residual insulin secretion in adults with autoimmune diabetes. Diabetes 1996; 45:1427–30.
- 99 Ortqvist E, Bjork E, Wallensteen M et al. Temporary preservation of beta-cell function by diazoxide treatment in childhood type 1 diabetes. Diabetes Care 2004; 27:2191–7.
- 100 Brod SA, Atkinson M, Lavis VR et al. Ingested IFN-alpha preserves residual beta cell function in type 1 diabetes. J Interferon Cytokine Res 2001; 21:1021–30.
- 101 Kempf K, Manzo G, Hanifi-Moghaddam P et al. Effect of combined oral proteases and flavonoid treatment in subjects at risk of Type 1 diabetes. Diabet Med 2009; 26:1309–10.
- 102 Herold KC, Gitelman SE, Willi SM et al. Teplizumab treatment may improve C-peptide responses in participants with type 1 diabetes after the new-onset period: a randomised controlled trial. Diabetologia 2013; 56:391–400.
- 103 Haller MJ, Wasserfall CH, Hulme MA et al. Autologous umbilical cord blood transfusion in young children with type 1 diabetes fails to preserve C-peptide. Diabetes Care 2011; 34:2567–9.
- 104 Haller MJ, Wasserfall CH, McGrail KM et al. Autologous umbilical cord blood transfusion in very young children with type 1 diabetes. Diabetes Care 2009; 32:2041–6.
- 105 Saudek F, Havrdova T, Boucek P, Karasova L, Novota P, Skibova J. Polyclonal anti-T-cell therapy for type 1 diabetes mellitus of recent onset. Rev Diabet Stud 2004; 1:80–8.
- 106 Giannoukakis N, Phillips B, Finegold D, Harnaha J, Trucco M. Phase I (safety) study of autologous tolerogenic dendritic cells in type 1 diabetic patients. Diabetes Care 2011; 34:2026–32.
- 107 Sumpter KM, Adhikari S, Grishman EK, White PC. Preliminary studies related to anti-interleukin-1beta therapy in children with newly diagnosed type 1 diabetes. Pediatr Diabetes 2011; 12:656– 67.
- 108 Mandrup-Poulsen T. IL-1 receptor antagonist in recent onset type 1 diabetes- a multicentre randomized, placedbo controlled trial [IT-SYO1]. Presented at: 72nd Scientific Session of the American Diabetes Association; 11 June 2012; Philadephia, PA.
- 109 Moran A. Canakinumab, an anti-IL-1 monoclonal antibody in recent -onset type 1 diabetes [IT-SY01]. Presented at: 72nd Scientific Session of the American Diabetes Association; 11 June 2012; Philadelphia, PA.
- 110 Walter M, Kaupper T, Adler K, Foersch J, Bonifacio E, Ziegler AG. No effect of the 1alpha,25-dihydroxyvitamin D3 on beta-cell residual function and insulin requirement in adults with newonset type 1 diabetes. Diabetes Care 2010; 33:1443–8.
- 111 Gu W, Hu J, Wang W *et al.* Diabetic ketoacidosis at diagnosis influences complete remission after treatment with hematopoietic stem cell transplantation in adolescents with type 1 diabetes. Diabetes Care 2012; **35**:1413–9.
- 112 Strom A, Kolb H, Martin S *et al.* Improved preservation of residual beta cell function by atorvastatin in patients with recent onset

- type 1 diabetes and high CRP levels (DIATOR trial). PLoS ONE 2012; 7:e33108.
- 113 Martin S, Herder C, Schloot NC et al. Residual beta cell function in newly diagnosed type 1 diabetes after treatment with atorvastatin: the Randomized DIATOR Trial. PLoS ONE 2011; 6:e17554.
- 114 Rother KI, Brown RJ, Morales MM *et al.* Effect of ingested interferon-alpha on beta-cell function in children with new-onset type 1 diabetes. Diabetes Care 2009; **32**:1250–5.
- 115 Faustman DL, Wang L, Okubo Y et al. Proof-of-concept, rand-omized, controlled clinical trial of bacillus-Calmette–Guérin for treatment of long-term type 1 diabetes. PLoS ONE 2012; 7:e41756.
- 116 Gottlieb PA, Quinlan S, Krause-Steinrauf H et al. Failure to preserve beta-cell function with mycophenolate mofetil and daclizumab combined therapy in patients with new-onset type 1 diabetes. Diabetes Care 2010; 33:826–32.

- 117 Couri CE, Oliveira MC, Stracieri AB et al. C-peptide levels and insulin independence following autologous nonmyeloablative hematopoietic stem cell transplantation in newly diagnosed type 1 diabetes mellitus. JAMA 2009; 301:1573–9.
- 118 Voltarelli JC, Couri CE, Stracieri AB et al. Autologous nonmyeloablative hematopoietic stem cell transplantation in newly diagnosed type 1 diabetes mellitus. JAMA 2007; 297:1568–76.
- 119 Barcala Tabarrozzi AE, Castro CN, Dewey RA, Sogayar MC, Labriola L, Perone MJ. Cell-based interventions to halt autoimmunity in type 1 diabetes mellitus. Clin Exp Immunol 2013; 171:135–46.
- 120 de Oliveira GL, Malmegrim KC, Ferreira AF *et al.* Up-regulation of fas and fasL pro-apoptotic genes expression in type 1 diabetes patients after autologous haematopoietic stem cell transplantation. Clin Exp Immunol 2012; **168**:291–302.